

EDITORIAL

Radiation resistance in breast cancer: are CD44⁺/CD24⁻/proteasome^{low}/PKH26⁺ cells to blame?

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See related research by Lagadec *et al.*, <http://breast-cancer-research.com/content/12/1/R13>

Abstract

Identification and characterization of cancer-initiating cells (CICs) enriched for stem cell-like functions and the establishment of a link between CICs and tumor recurrence, chemotherapy resistance and radiation resistance, and metastasis have been the focus of cancer research for the last eight years. Although this field has its share of controversies, it is becoming apparent that cells isolated from recurrent or residual tumors or both are enriched for cancer cells that have a specific phenotype compared with heterogeneous cells in the primary tumor. Enrichment of CICs in tumors subjected to radiation therapy could be due in part to the delivery of sublethal doses of treatment and the efficient radical scavenging system within CICs. Sublethal doses of radiation are sufficient to induce senescence of non-CICs while forcing CICs to gain several new properties related to cell cycle progression in addition to maintaining or enhancing stem cell characteristics of pre-treatment CICs. Characterizing pathways responsible for the increase in CICs after therapy and exploiting the unique characteristics of therapy-resistant CICs for developing targeted therapies are becoming a central focus of research in the rapidly evolving field of CICs.

In the previous issue of *Breast Cancer Research*, Lagadec and colleagues [1] used breast cancer cell line models to examine the self-renewal and cell cycle properties of cancer cells that survive during the course of fractionated sublethal radiation. Previous studies from the same group and others have demonstrated enrichment or resistance

of cancer-initiating cells (CICs) during the course of fractionated radiation [2,3]. The current report demonstrates that the enriched CICs are capable of enhanced self-renewal for three generations compared with CICs from non-irradiated cells. Contrary to non-irradiated CICs, which are non-proliferating and are predominantly in the G₀ phase of the cell cycle, radiation mobilizes CICs from a quiescent to a proliferative state (G₂ phase). Whether CICs undergo similar cell cycle changes *in vivo* upon radiation and whether cycling CICs become susceptible to drugs targeting cells moving through G₂ phase remain to be determined.

Cancer cells expressing the cell surface marker CD44 but not CD24 (CD44⁺/CD24^{-/low}) were the first described breast cancer CICs [4]. Subsequent studies have identified additional phenotypic or functional markers or both to characterize CICs: elevated levels of aldehyde dehydrogenase (ALDH1⁺) [5], lower levels of proteasome activity [6], enhanced PKH26 dye-retaining capacity [7], CD24^{high}/CD49F^{high}/Delta-notch-like EGF (epidermal growth factor) repeat-containing transmembrane (DNER)^{high} [7], CD24^{high}/CD49F^{high}/Delta-like 1 (DLL1)^{high} [7], CD49F⁺/DLL1^{high}/DNER^{high}, and expression of a mammosphere signature, including low expression of claudin [8]. CD44⁺/CD24⁻ CICs and non-CD44⁺/CD24⁻ cells expressing different levels of these markers have been tested in *in vitro* (mammosphere) and *in vivo* (tumorigenicity in NOD/SCID [non-obese diabetic/severe combined immunodeficiency disease] mice) assays. CD44⁺/CD24⁻/ALDH1⁺ cells display the highest level of CIC activity in tumorigenicity assays compared with CD44⁺/CD24⁻ or ALDH1⁺ cells [5]. Lagadec and colleagues [1] observed a subpopulation of CD44⁺/CD24^{-/low} cells with low proteasome activity, although this specific subpopulation was not characterized further for stemness. Although these studies are beginning to refine CICs into distinct subgroups, the remaining question is which of the phenotypically distinct CICs are responsible for chemotherapy resistance and radiation resistance and organ-specific metastasis. Also, it is

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unclear whether the above markers can identify CICs for all intrinsic subtypes of breast cancer [9] or whether these markers favor characterization of CICs derived from the basal subtype or the HER2⁺ subtype of breast cancer [10,11].

CD44⁺/CD24^{-/low} cells are generally enriched in basal subtype breast cancers as well as in cell lines that have undergone epithelial-to-mesenchymal transition [10,12]. In contrast, luminal type breast cancers that express estrogen receptor-alpha (ER α) contain less than 1% CD44⁺/CD24^{-/low} cells [10]. Lagadec and colleagues [1] were able to demonstrate enrichment of CD44⁺/CD24⁻ cells upon fractionated radiation of ER α ⁺ breast cancer cell lines MCF-7 and T47-D. A significant number of enriched cells retained PKH26 dye and contained low proteasome activity compared with non-irradiated cells or cells with high proteasome activity. Thus, radiation-resistant cells display three characteristics of CICs: CD44⁺/CD24^{-/low}, low proteasome, and PKH26⁺. Interestingly, enriched cells from T47-D cells, but not from MCF-7 cells, displayed enhanced self-renewal capacity for three generations, suggesting differences in CICs between ER α breast cancer cell lines. Both MCF-7 and T47-D cells represent luminal type A breast cancer, which generally belongs to a favorable prognostic subgroup [13]. However, the two cell lines differ in their p53 mutation status; T47-D cells contain mutant p53, whereas p53 is wild-type in the MCF-7 cells [14]. Based on the previously described role of p53 in cancer stem cells in mouse models [15], it is tempting to speculate that p53 status is one of the factors that determine the self-renewal capacity of cancer cells that survive a sublethal dose of radiation. It is important to emphasize that the enrichment of CICs was observed only upon exposure to a sublethal dose of radiation. Therefore, similar enrichment of CICs may not occur in the clinical setting unless a significant fraction of cancer cells receive a sublethal dose or contain a p53 mutation or both. In this respect, recent clinical trials have demonstrated significant survival benefits of radiation therapy for breast cancer patients [16]. Further optimization of radiation delivery to avoid exposure to sublethal doses may lead to increased benefits of radiation therapy.

Abbreviations

ALDH1, aldehyde dehydrogenase; CIC, cancer-initiating cell; DLL1, Delta-like 1; DNER, Delta-notch-like EGF (epidermal growth factor) repeat-containing transmembrane; ER α , estrogen receptor-alpha.

Competing interests

The author declares that he has no competing interests.

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References

1. Lagadec C, Vlashi E, Della Donna L, Meng Y, Dekmezian C, Kim K, Pajonk F: Survival, self-renewing capacity and multi-lineage potency of breast

- cancer initiating cells during fractionated radiation treatment. *Breast Cancer Res* 2010, **12**:R13.
2. Diehn M, Cho RW, Lobo NA, Kalisky T, Dorie MJ, Kulp AN, Qian D, Lam JS, Ailles LE, Wong M, Joshua B, Kaplan MJ, Wapnir I, Dirbas FM, Somlo G, Garberoglio C, Paz B, Shen J, Lau SK, Quake SR, Brown JM, Weissman IL, Clarke MF: Association of reactive oxygen species levels and radioresistance in cancer stem cells. *Nature* 2009, **458**:780-783.
3. Phillips TM, McBride WH, Pajonk F: The response of CD24^(-low)/CD44⁺ breast cancer-initiating cells to radiation. *J Natl Cancer Inst* 2006, **98**:1777-1785.
4. Al-Hajj M, Wicha MS, Benito-Hernandez A, Morrison SJ, Clarke MF: Prospective identification of tumorigenic breast cancer cells. *Proc Natl Acad Sci U S A* 2003, **100**:3983-3988.
5. Ginestier C, Hur MH, Charafe-Jauffret E, Monville F, Dutcher J, Brown M, Jacquemier J, Viens P, Kleer CG, Liu S, Schott A, Hayes D, Birnbaum D, Wicha MS, Dontu G: ALDH1 is a marker of normal and malignant human mammary stem cells and a predictor of poor clinical outcome. *Cell Stem Cell* 2007, **1**:555-567.
6. Vlashi E, Kim K, Lagadec C, Donna LD, McDonald JT, Eghbali M, Sayre JW, Stefani E, McBride W, Pajonk F: *In vivo* imaging, tracking, and targeting of cancer stem cells. *J Natl Cancer Inst* 2009, **101**:350-359.
7. Pece S, Tosoni D, Confalonieri S, Mazzarol G, Vecchi M, Ronzoni S, Bernard L, Viale G, Pelicci PG, Di Fiore PP: Biological and molecular heterogeneity of breast cancers correlates with their cancer stem cell content. *Cell* 2010, **140**:62-73.
8. Creighton CJ, Li X, Landis M, Dixon JM, Neumeister VM, Sjolund A, Rimm DL, Wong H, Rodriguez A, Herschkowitz JI, Fan C, Zhang X, He X, Pavlick A, Gutierrez MC, Renshaw L, Larionov AA, Faratian D, Hilsenbeck SG, Perou CM, Lewis MT, Rosen JM, Chang JC: Residual breast cancers after conventional therapy display mesenchymal as well as tumor-initiating features. *Proc Natl Acad Sci U S A* 2009, **106**:13820-13825.
9. Sørlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, van de Rijn M, Jeffrey SS, Thorsen T, Quist H, Matese JC, Brown PO, Botstein D, Eystein Lønning P, Børresen-Dale AL: Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci U S A* 2001, **98**:10869-10874.
10. Sheridan C, Kishimoto H, Fuchs RK, Mehrotra S, Bhat-Nakshatri P, Turner CH, Goulet R Jr, Badve S, Nakshatri H: CD44⁺/CD24⁻ breast cancer cells exhibit enhanced invasive properties: an early step necessary for metastasis. *Breast Cancer Res* 2006, **8**:R59.
11. Park SY, Lee HE, Li H, Shipitsin M, Gelman R, Polyak K: Heterogeneity for stem cell-related markers according to tumor subtype and histologic stage in breast cancer. *Clin Cancer Res* 2010, **16**:876-887.
12. Mani SA, Guo W, Liao MJ, Eaton EN, Ayyanan A, Zhou AY, Brooks M, Reinhard F, Zhang CC, Shipitsin M, Campbell LL, Polyak K, Brisken C, Yang J, Weinberg RA: The epithelial-mesenchymal transition generates cells with properties of stem cells. *Cell* 2008, **133**:704-715.
13. Neve RM, Chin K, Fridlyand J, Yeh J, Baehner FL, Fevr T, Clark L, Bayani N, Coppe JP, Tong F, Speed T, Spellman PT, DeVries S, Lapuk A, Wang NJ, Kuo WL, Stiwell JL, Pinkel D, Albertson DG, Waldman FM, McCormick F, Dickson RB, Johnson MD, Lippman M, Ethier S, Gazdar A, Gray JW: A collection of breast cancer cell lines for the study of functionally distinct cancer subtypes. *Cancer Cell* 2006, **10**:515-527.
14. Casey G, Lo-Hsueh M, Lopez ME, Vogelstein B, Stanbridge EJ: Growth suppression of human breast cancer cells by the introduction of a wild-type p53 gene. *Oncogene* 1991, **6**:1791-1797.
15. Zhang M, Behbod F, Atkinson RL, Landis MD, Kittrell F, Edwards D, Medina D, Tsimelzon A, Hilsenbeck S, Green JE, Michalowska AM, Rosen JM: Identification of tumor-initiating cells in a p53-null mouse model of breast cancer. *Cancer Res* 2008, **68**:4674-4682.
16. Darby S: Overview of the randomized trials of radiotherapy in early breast cancer (abstract MS 3-1). *Cancer Res* 2009, **69**:486S.

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