Short communication **Adjuvant treatment: the contribution of expression microarrays**

Per Eystein Lønning^{1,2}, Ranjan Chrisanthar^{1,3}, Vidar Staalesen^{1,3}, Stian Knappskog^{1,3} and Johan Lillehaug³

¹Section of Oncology, Institute of Medicine, University of Bergen, Bergen N-5020, Norway ²Department of Oncology, Haukeland University Hospital, Bergen N-5020, Norway ³Department of Molecular Biology, University of Bergen, Bergen N-5020, Norway

Corresponding author: Per Eystein Lønning, per.lonning@helse-bergen.no

Published: 20 December 2007 This article is online at http://breast-cancer-research.com/content/9/S2/S14 © 2007 BioMed Central Ltd Breast Cancer Research 2007, 9(Suppl 2):S14 (doi:10.1186/bcr1812)

Introduction

Although gene expression microarrays provide novel tools and hold great promise in cancer research, achievements thus far in terms of improved prognostication and, in particular, prediction of drug sensitivity have been moderate. To improve clinical therapy, we believe that it is imperative to integrate gene expression arrays with other laboratory methods based on functional concepts [1,2].

Breast cancer taxonomy

The first study to explore human breast cancer biology applying gene expression signatures was that reported by Perou and coworkers [3] in 2000. Here, oestrogen receptor (ER)-positive breast cancers (designated luminal class, based on cytokeratin expression) were found to be associated with particular gene expression profiles. Moreover, the gene expression signatures revealed (at least) two distinct subclasses among the ER-positive tumours, termed luminal A and luminal B. This subclassification provided novel prognostic information. Thus, among patients with locally advanced breast cancers undergoing primary chemotherapy with either doxorubicin monotherapy [4] or 5-fluorouracil and mitomycin given in concert [5] to be followed by tamoxifen adjuvant for 5 years, a poor prognosis was identified among patients with tumours expressing a luminal B profile as opposed to the luminal A group [6]. Interestingly, when the classification was applied to a second cohort of patients with early stage breast cancers who had not received adjuvant endocrine therapy [7], again the luminal A and B classes were associated with different prognosis; the relative difference, however, was much less than that in patients receiving tamoxifen treatment. Although this could indicate a predictive component (higher sensitivity for luminal A class tumours to tamoxifen treatment compared with luminal B ones), such conclusions should not be inferred from indirect comparison.

The second major achievement was further subclassification within the group of ER-negative tumours. This led to identification of the so-called 'triple negative' class (tumours negative with respect to expression of ER and progesterone receptor that, in addition, lack over-expression and/or amplification of HER2) as a distinct subclass. These triple negative tumours expressed keratin markers that are strongly suggestive of a basal cell origin (for which reason they are frequently referred to as 'basal cell class' tumours), contrasting with the luminal origin of breast cancers in general.

Prognostication

Subsequently, several studies [8-13] have identified different gene expression profiles as being associated with prognostication in breast cancer [8-13]. Notably, however, the various profiles identified differ considerably with respect to genes included, and the extraction of multiple signatures from the same dataset questions the specificity of such signatures [14]. Others have argued that the information provided may not be superior to what is achieved by optimal use of conventional factors [15]. Moreover, because these studies in general were conducted retrospectively in unselected patient cohorts, meaning that the patients were exposed to various drug regimens, the issue of potentially predictive components may not be excluded. Notably, a main reason why lymph node status may be used as a single marker to select high-risk patients for adjuvant therapy based on risk for having a relapse is due to the fact that it is a 'pure' prognostic factor; patients defined as having a poor prognosis are not more likely to be therapy resistant than those having a better prognosis. This underlines a general principle. When looking for novel prognostic factors, it is mandatory to keep in mind that no prognostic factor may be defined and implemented for clinical use without detailed knowledge regarding its potential predictive effect for the therapy applied [16,17]. Considering a factor such as TP53, mutations that affect the DNA-binding part of the protein are associated with a poor

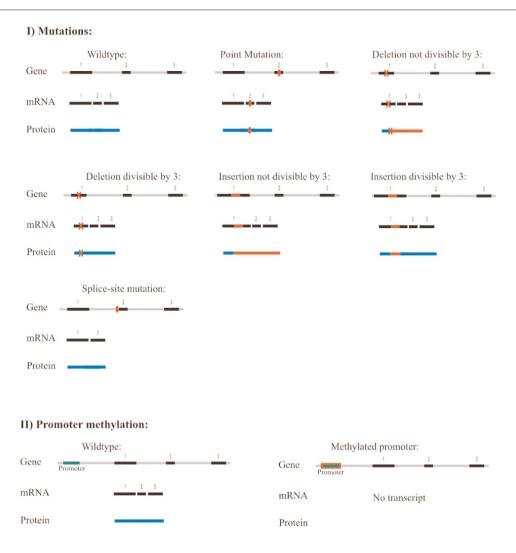


Figure 1

Different types of gene mutations and mechanisms of epigenetic silencing. Reproduced with permission from [16].

prognosis [18]; however, they also confer poor sensitivity to chemotherapeutics such as anthracyclines and mitomycin [5,19].

Predicting response to therapy

Primary medical therapy (previously termed 'neoadjuvant treatment') represents an optimal setting in which to study drug effects on tumours directly [20]. Thus, several studies have explored gene expression profiles that predict responsiveness to different chemotherapeutic regimens, including taxane monotherapy [21,22] or anthracycline- or mitomycin-containing regimens administered as monotherapy [23] or in combination with other drugs [23-25], including taxanes [24,26-29]. The general conclusion from these studies may be summarized as follows. First, independent of regimen and the statistical approach (supervised or unsupervised), there is in general a correlation between gene expres-

sion profiles and responsiveness to therapy. Second, for none of the signatures identified has the combined sensitivity and specificity reached a level that allows its implementation for clinical use outside trials. Third, in none of these signatures has the value repeatedly been corroborated by other investigators.

Interestingly, looking at response to therapy across the different breast cancer subclasses [23,25], some differences in responsiveness could be detected. However, these differences were not of sufficient magnitude to allow clinical application to therapy selection.

There are several limitations to the use of microarray analysis as a single method for exploring tumour biology. The spectrum of pathological events that lead to disturbed gene function is huge [16], involving components such as large

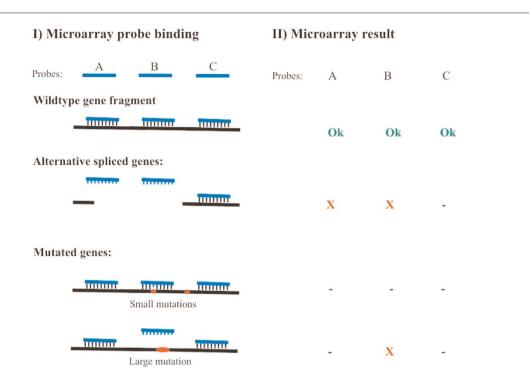


Figure 2

Microarray signals are generated by hybridizing a probe to the gene product (complementary DNA [cDNA]). Alternative splicing, as well as different types of mutations, may influence the result but may go undetected as well, depending on the exact type of lesion as well as its location with respect to the area hybridising with the probe. 'X' and '-' indicate potential errors; 'X' indicates wrongly detected transcript levels, while '-' signifies correct levels reported, but with other undetectable lesions present in the gene. Reproduced with permission from [16].

and small deletions or single base substitutions, mutations that affect promoter regions or splice-sites, as well as epigenetic silencing (Figure 1). In addition, the issue of multiple splice variants generated from the same gene has received increasing attention [30]. Alternative splices may be transcribed into protein products with different biological function [31]; whether such splices are detected together with the main transcript on microarrays depends on whether the sequence covered by the probe is included in the splice transcript and how the mutation affects hybridization (Figure 2). Additionally, the stability of the different splice transcripts and encoded proteins may vary considerably. Finally, many signalling pathways, including activation of p53 [32], involve post-translational modifications such as protein phosphorylations, deacetylations, and so on, meaning that information relevant to changes in biological function of specific proteins is not reflected in altered mRNA expression.

It is clear (Figures 1 and 2) that although microarrays may provide information about transcriptional status of individual genes, interactions such as inclusion of alternative splices may confound the biological interpretation. Mutations that affect genes encoding proteins that are involved upstream or downstream in a particular functional cascade may generate different overall gene expression profiles, despite having similar effects on this particular pathway [2]. On the other hand, a single mutation in a critical gene may have profound biological effects despite having a limited effect on the total gene expression profile. Thus, to identify defects in functional pathways that lead to outcomes such as drug resistance, we need a panel of methods that detects different pathological disturbances based on functional hypotheses [2].

Adjuvant therapy

In contrast to the number of studies conducted in the primary medical setting, data with respect to the predictive value of gene expression profiles in the adjuvant setting are scarce. Although studies have addressed genetic factors that determine prognosis in patient cohorts exposed to defined therapies such as tamoxifen [33], the only large study exploring gene expression profile with respect to benefit of chemotherapy was that conducted by the NSABP (National Surgical Adjuvant Breast and Bowel Project). Taking a 21gene expression signature previously shown to be associated with prognosis in tamoxifen-treated breast cancer patients [34], they reported that the same profiles also predict the likelihood of benefit from adjuvant chemotherapy with a regimen containing cyclophosphamide, methotrexate, and 5fluorouracil [35]. Notably, they found a high recurrence score to be associated with profound effect of chemotherapy; this is in contrast to the intermediate and low scores, for which no significant clinical benefit of chemotherapy was achieved. Although this test has been implemented in many centres around the world, independent validation is still awaited.

Testing for chemo-resistance *in vivo*: adjuvant versus primary medical treatment as the optimal setting

Although primary medical therapy is considered to be the optimal way to assess direct antitumour efficacy of drug treatment, this may not automatically imply a correlation with outcome defined as general relapse or cancer death. Pathological complete response to primary medical treatment has clearly been correlated with long-term prognosis [27]; however, several patients achieving a complete response may later relapse. There may be a number of explanations for this observation, such as survival of resistant subclones among micrometastases. However, we should recognize that a number of biological parameters in addition to direct drug sensitivity are involved in the metastatic process, such as blood vessel wall invasion, tumour-host organ interactions and angiogenesis. Notably, gene signatures have been identified that predict organ-specific metastatic propensity in experimental as well as clinical materials [36-38]. Interestingly, Massagué and coworkers [39], in addition, have identified a few key genes from their lung metastases signature associated with growth, invasion and angiogenesis, which play a key role in regulating lung metastases in experimental systems. Although adjuvant studies need larger patient cohorts as well as longer follow up in comparison with studies of primary medical therapy, there is clearly a need for long-term follow up of patients undergoing primary medical as well as adjuvant therapy to address these issues.

Acknowledgement

This article has been published as part of Breast Cancer Research Volume 9 Supplement 2, 2007: Controversies in Breast Cancer. The full contents of the supplement are available online at http://breast-cancer-research.com/supplements/9/S2.

References

- Lønning PE, Sørlie T, Børresen-Dale A-L: Genomics in breast cancer - therapeutic implications? Nat Clin Pract Oncol 2005, 2:26-33.
- Lønning PE: Genes causing inherited cancer as beacons identifying the mechanisms of chemoresistance. *Trends Mol Med* 2004, 10:113-118.
- Perou CM, Sørlie T, Eisen MB, van de Rijn M, Jeffrey SS, Rees CA, Pollack JR, Ross DT, Johnsen H, Akslen LA, et al.: Molecular portraits of human breast tumours. Nature 2000, 406:747-752.
- Geisler S, Lønning PE, Aas T, Johnsen H, Fluge O, Haugen DF, Lillehaug JR, Akslen LA, Børresen-Dale AL: Influence of TP53 gene alterations and c-erbB-2 expression on the response to treatment with doxorubicin in locally advanced breast cancer. *Cancer Res* 2001, 61:2505-2512.
- Geisler S, Børresen-Dale AL, Johnsen H, Aas T, Geisler J, Akslen LA, Anker G, Lønning PE: TP53 gene mutations predict the response to neoadjuvant treatment with FUMI in locally advanced breast cancer. *Clin Cancer Res* 2003, 9:5582-5588.
- Sørlie T, Perou CM, Tibshirani R, Aas T, Geisler S, Johnsen H, Hastie T, Eisen MB, van de Rijn M, Jeffrey SS, *et al.*: Gene expression patterns of breast carcinomas distinguish tumor subclasses with clinical implications. *Proc Natl Acad Sci USA*

2001, **98:**10869-10874.

- Sørlie T, Tibshirani R, Parker J, Hastie T, Marron JS, Nobel A, Deng S, Johnsen H, Pesich R, Geisler S, et al.: Repeated observation of breast tumor subtypes in independent gene expression data sets. Proc Natl Acad Sci USA 2003, 100:8418-8423.
- van de Vijver MJ, He YD, van't Veer LJ, Dai H, Hart AA, Voskuil DW, Schreiber GJ, Peterse JL, Roberts C, Marton MJ, et al.: A gene-expression signature as a predictor of survival in breast cancer. N Engl J Med 2002, 347:1999-2009.
- Sotiriou C, Neo SY, McShane LM, Korn EL, Long PM, Jazaeri A, Martiat P, Fox SB, Harris AL, Liu ET: Breast cancer classification and prognosis based on gene expression profiles from a population-based study. *Proc Natl Acad Sci USA* 2003, 100: 10393-10398.
- Ahr A, Karn T, Solbach C, Seiter T, Strebhardt K, Holtrich U, Kaufmann M: Identification of high risk breast-cancer patients by gene expression profiling. *Lancet* 2002, 359:131-132.
- Huang E, Cheng SH, Dressman H, Pittman J, Tsou MH, Horng CF, Bild A, Iversen ES, Liao M, Chen CM, et al.: Gene expression predictors of breast cancer outcomes. Lancet 2003, 361: 1590-1596.
- Onda M, Emi M, Nagai H, Nagahata T, Tsumagari K, Fujimoto T, Akiyama F, Sakamoto G, Makita M, Kasumi F, *et al.*: Gene expression patterns as marker for 5-year postoperative prognosis of primary breast cancer. J Cancer Res Clin Oncol 2004, 130:537-545.
- Wang Y, Klijn JG, Zhang Y, Sieuwerts AM, Look MP, Yang F, Talantov D, Timmermans M, Meijer-van Gelder ME, Yu J, et al.: Gene-expression profiles to predict distant metastasis of lymph-node-negative primary breast cancer. Lancet 2005, 365:671-679.
- Ein-Dor L, Kela I, Getz G, Givol D, Domany E: Outcome signature genes in breast cancer: is there a unique set? *Bioinformatics* 2005, 21:171-178.
- Edén P, Ritz C, Rose C, Fernö M, Peterson C: 'Good Old' clinical markers have similar power in breast cancer prognosis as microarray gene expression profilers. *Eur J Cancer* 2004, 40: 1837-1841.
- Lønning PE, Knappskog S, Staalesen V, Chrisanthar R, Lillehaug JR: Breast cancer prognostication and prediction in the postgenomic era. Ann Oncol 2007, 18:1293-1306.
- Lønning PE: Breast cancer prognostication and prediction; are we making progress? Ann Oncol 2007, 18(Suppl 8):viii3-viii7.
- Børresen AL, Andersen TI, Eyfjörd JE, Cornelis RS, Thorlacius S, Borg A, Johansson U, Theillet C, Scherneck S, Hartman S: TP53 mutations and breast cancer prognosis: Particularly poor survival rates for cases with mutations in the zinc-binding domains. Genes Chromosomes Cancer 1995, 14:71-75.
- Aas T, Børresen AL, Geisler S, Smith-Sørensen B, Johnsen H, Varhaug JE, Akslen LA, Lønning PE: Specific P53 mutations are associated with *de novo* resistance to doxorubicin in breast cancer patients. *Nature Med* 1996, 2:811-814.
- 20. Lønning P: Study of suboptimum treatment response: lessons from breast cancer. Lancet Oncol 2003, 4:177-185.
- Chang JC, Wooten EC, Tsimelzon A, Hilsenbeck SG, Gutierrez MC, Elledge R, Mohsin S, Osborne CK, Chamness GC, Allred DC, et al.: Gene expression profiling for the prediction of therapeutic response to docetaxel in patients with breast cancer. *Lancet* 2003, 362:362-369.
- Iwao-Koizumi K, Matoba R, Ueno N, Kim SJ, Ando A, Miyoshi Y, Maeda E, Noguchi S, Kato K: Prediction of docetaxel response in human breast cancer by gene expression profiling. J Clin Oncol 2005, 23:422-431.
- Sørlie T, Perou CM, Fan C, Geisler S, Aas T, Nobel A, Anker G, Akslen LA, Botstein D, Børresen-Dale AL, Lønning PE: Gene expression profiles do not consistently predict the clinical treatment response in locally advanced breast cancer. *Mol Cancer Ther* 2006, 5:2914-2918.
- 24. Hannemann J, Oosterkamp HM, Bosch CA, Velds A, Wessels LF, Loo C, Rutgers EJ, Rodenhuis S, van de Vijver MJ: Changes in gene expression associated with response to neoadjuvant chemotherapy in breast cancer. *J Clin Oncol* 2005, 23:3331-3342.
- Rouzier R, Perou CM, Symmans WF, Ibrahim N, Cristofanilli M, Anderson K, Hess KR, Stec J, Ayers M, Wagner P, *et al.*: Breast cancer molecular subtypes respond differently to preoperative chemotherapy. *Clin Cancer Res* 2005, 11:5678-5685.

- Ayers M, Symmans WF, Stec J, Damokosh AI, Clark E, Hess K, Lecocke M, Metivier J, Booser D, Ibrahim N, et al.: Gene expression profiles predict complete pathologic response to neoadjuvant paclitaxel and fluorouracil, doxorubicin, and cyclophosphamide chemotherapy in breast cancer. J Clin Oncol 2004, 22:2284-2293.
- Gianni L, Zambetti M, Clark K, Baker J, Cronin M, Wu J, Mariani G, Rodriguez J, Carcangiu M, Watson D, et al.: Gene expression profiles in paraffin-embedded core biopsy tissue predict response to chemotherapy in women with locally advanced breast cancer. J Clin Oncol 2005, 23:7265-7277.
- Thuerigen O, Schneeweiss A, Toedt G, Warnat P, Hahn M, Kramer H, Brors B, Rudlowski C, Benner A, Schuetz F, et al.: Gene expression signature predicting pathologic complete response with gemcitabine, epirubicin, and docetaxel in primary breast cancer. J Clin Oncol 2006, 24:1839-1845.
- Dressman HK, Hans C, Bild A, Olson JA, Rosen E, Marcom PK, Liotcheva VB, Jones EL, Vujaskovic Z, Marks J, et al.: Gene expression profiles of multiple breast cancer phenotypes and response to neoadjuvant chemotherapy. Clin Cancer Res 2006, 12:819-826.
- Staalesen V, Falck J, Geisler S, Bartkova J, Børresen-Dale AL, Lukas J, Lillehaug JR, Bartek J, Lønning PE: Alternative splicing and mutation status of CHEK2 in stage III breast cancer. Oncogene 2004, 23:8535-8544.
- 31. Stiewe T, Putzer BM: p73 in apoptosis. Apoptosis 2001, 6:447-452.
- Liu G, Chen XB: Regulation of the p53 transcriptional activity. J Cell Biochem 2006, 97:448-458.

- Ma XJ, Wang Z, Ryan PD, Isakoff SJ, Barmettler A, Fuller A, Muir B, Mohapatra G, Salunga R, Tuggle JT, et al.: A two-gene expression ratio predicts clinical outcome in breast cancer patients treated with tamoxifen. Cancer Cell 2004, 5:607-616.
- Paik S, Shak S, Tang G, Kim C, Baker J, Cronin M, Baehner FL, Walker MG, Watson D, Park T, et al.: A multigene assay to predict recurrence of tamoxifen-treated, node-negative breast cancer. N Engl J Med 2004, 351:2817-2826.
- Paik S, Tang G, Shak S, Kim C, Baker J, Kim W, Cronin M, Baehner FL, Watson D, Bryant J, et al.: Gene expression and benefit of chemotherapy in women with node-negative, estrogen receptor-positive breast cancer. J Clin Oncol 2006, 24:3726-3734.
- Kang Y, He W, Tulley S, Gupta GP, Serganova I, Chen CR, Manova-Todorova K, Blasberg R, Gerald WL, Massagué J: Breast cancer bone metastasis mediated by the Smad tumor suppressor pathway. Proc Natl Acad Sci USA 2005, 102:13909-13914.
- Minn AJ, Gupta GP, Siegel PM, Bos PD, Shu W, Giri DD, Viale A, Olshen AB, Gerald WL, Massagué J: Genes that mediate breast cancer metastasis to lung. *Nature* 2005, 436:518-524.
 Smid M, Wang Y, Klijn JG, Sieuwerts AM, Zhang Y, Atkins D,
- Smid M, Wang Y, Klijn JG, Sieuwerts AM, Zhang Y, Atkins D, Martens JW, Foekens JA: Genes associated with breast cancer metastatic to bone. J Clin Oncol 2006, 24:2261-2267.
- Gupta GP, Nguyen DX, Chiang AC, Bos PD, Kim JY, Nadal C, Gomis RR, Manova-Todorova K, Massagué J: Mediators of vascular remodelling co-opted for sequential steps in lung metastasis. *Nature* 2007, 446:765-770.