### Review

## Direct effects of bisphosphonates on breast cancer cells

Siddhika G Senaratne and Kay W Colston

Department of Oncology, Gastroenterology, Endocrinology, and Metabolism, St George's Hospital Medical School, Cranmer Terrace, London, UK

Correspondence: Kay W Colston, PhD, Department of Oncology, Gastroenterology, Endocrinology and Metabolism, St George's Hospital Medical School, Cranmer Terrace, London SW17 0RE, UK. Tel: +44 020 8725 5887; fax: +44 020 8725 0513; e-mail: k.colston@sqhms.ac.uk

Received: 3 September 2001 Breast Cancer Res 2002, 4:18-23

Revisions requested: 25 September 2001 Revisions received: 3 October 2001

Accepted: 9 October 2001 © 2002 BioMed Central Ltd

Published: 14 November 2001 (Print ISSN 1465-5411; Online ISSN 1465-542X)

#### **Abstract**

In addition to inhibiting bone resorption, bisphosphonates have also been shown to exhibit antitumour effects. *In vitro*, bisphosphonates inhibit proliferation and induce apoptosis in cultured human breast cancer cells. In addition, bisphosphonate treatment interferes with breast cancer cell adhesion to bone matrix, and inhibits cell migration and invasion. The combination of bisphosphonates with other anticancer drugs such as the taxoids markedly enhances these effects. These newly recognized direct actions of bisphosphonates on breast cancer cells indicate that these agents may have a greater role to play in treatment of patients suffering from cancers with a propensity to metastasize to bone.

 $\textbf{Keywords:} \ a poptosis, \ b is phosphonates, \ breast \ cancer, \ cell \ adhesion, \ invasion$ 

### Introduction

Over 80% of women with advanced breast cancer ultimately develop bone metastases that result in significant morbidity and mortality. Breast cancer metastases in bone can cause intractable pain, bone fracture, spinal cord compression and hypercalcaemia [1–3]. From the moment breast cancer cells arrive in the bone microenvironment, however, they stimulate bone resorption with subsequent selective increase in the attraction and growth of new cancer cells to bone [4]. Therefore, any treatment aimed at palliation or perhaps even prevention of bone metastases should focus on disrupting this attraction and growth, which are involved in the initiation and amplification of the metastatic process.

Bisphosphonates are widely used for the treatment of bone metastases, and an increasing body of evidence suggests that these compounds provide benefit to breast cancer patients with secondary cancers in bone [5]. Bisphosphonates are analogues of endogenous pyrophosphates in which a carbon atom replaces the central atom of oxygen. *In vivo*, bisphosphonates bind strongly to

hydroxyapatite on the bone surface and are preferentially delivered to sites of increased bone formation or resorption. They are potent inhibitors of osteoclast-mediated bone resorption [6] and are effective in lowering serum calcium concentrations in patients with hypercalcaemia of malignancy [7,8]. Treatment with bisphosphonates has also been shown to reduce skeletal morbidity significantly and to improve quality of life in breast cancer patients with bone metastases [7].

The mechanisms by which bisphosphonates inhibit osteoclast-mediated bone resorption appear to involve an inhibition of formation of osteoclasts from immature precursor cells [6,9,10] or direct inhibition of resorption via induction of apoptosis in mature osteoclasts [9,11,12]. Furthermore, as outlined elsewhere, bisphosphonate treatment has been shown to inhibit the progression and development of bone metastases in a mouse model of breast cancer [13,14]. Such a beneficial effect of bisphosphonates on tumour burden in bone may result from a direct antitumour effect on breast cancer cells. Evidence is now emerging that this is indeed the case and that treatment of cultured human breast cancer cells treated with bisphosphonates induces inhibitory effects on adhesion, invasion and cell survival.

# Effects of bisphosphonates on cell adhesion to and invasion of bone

It is well accepted that adhesion of cancer cells to bone matrix is a vital step in the bone metastasis process, and it has been suggested that exposure of bone to bisphosphonates could alter properties of the bone matrix that are required for adhesion of breast cancer cells. In this regard, previous studies [6,9,15] have indicated that exposure of calcified matrix of bone to bisphosphonates *in vitro* alters the properties of the bone matrix that are required for attachment of the osteoclast.

Initially, the ability of human breast cancer cells to adhere to bone matrices that had been pretreated with bisphosphonates was investigated by van der Pluijm et al. [16]. In those experiments adhesion of MDA-MB-231 human breast cancer cells to bovine cortical bone slices and sections of developing trabecular bone from neonatal mouse tail were assessed. Those studies showed that pretreatment of bone matrices with certain bisphosphonates at concentrations of 1-100 µmol/l not only prevented adhesion of breast cancer cells to bone matrix, but also inhibited cell spreading. However, of the bisphosphonates tested only pretreatment of matrices with nitrogen-containing bisphosphonates (pamidronate, olpandronate, alendronate and ibandronate) led to these inhibitory effects. lbandronate was found to be the most potent compound. Pretreatment with clodronate or etidronate did not affect adhesion to bone matrix or cell spreading, and the order of potency of the six bisphosphonates corresponded to their ranking in bone resorption assays [6,9]. No effects on cell viability were observed over the 3 h period during which the cells were allowed to adhere to bone matrices.

Subsequently, Boissier et al. [17] evaluated the effect of direct treatment of breast cancer cells with bisphosphonates on their ability to adhere to unmineralized and mineralized bone extracellular matrices. Using mineralized bovine cortical bone slices and unmineralized extracellular matrices produced by cultured osteoblastic cells, those investifound that pretreatment of MCF-7 MDA-MB-231 breast cancer cells for 24 h with bisphosphonates inhibited cell adhesion. Similar effects were found with cultured prostatic carcinoma cells. Of the bisphosphonates used, only ibandronate, NE-10244 (antiresorptive active pyridinium analogue of risedronate) and pamidronate inhibited cell adhesion at low concentrations, with half-maximal inhibitions at 5 pmol/l, 0.1 nmol/l and 10 nmol/l, respectively. Clodronate achieved the same inhibitory effects at a high concentration, with half-maximal inhibition at 10 µmol/l. At concentrations that inhibited cell adhesion after 24 h of treatment, no effects of bisphosphonates on cell viability or integrin expression were detected.

Figure 1

$$\begin{array}{c|cccc}
O^{-} & R_{1} & O^{-} \\
 & | & | & | \\
O \Longrightarrow P \longrightarrow C \longrightarrow P \Longrightarrow O \\
 & | & | & | \\
O^{-} & R_{2} & O^{-}
\end{array}$$

Structure of bisphosphonates.

The mechanism by which the various bisphosphonates inhibit cell adhesion when coated onto mineralized or unmineralized matrix does not relate to their direct inhibitory action when incubated with cells *in vitro*. Thus, the analogue NE-58051 (inactive pyridylpropylidene analogue of risedronate, which lacks a methyl group in the R2 chain of the molecule; Fig. 1) was effective in preventing cell adhesion when cortical bone slices were coated with this compound [17]. However, pretreatment of cultured breast cancer cells *in vitro* with NE-58051, before seeding onto uncoated bone slices, did not affect adhesion.

The taxoids taxol and taxotere are effective antitumour compounds that are currently used routinely in the treatment of metastatic breast carcinoma. Magnetto et al. [18] determined whether there could be additive or synergistic effects of bisphosphonates in combination with taxoids on adhesion of breast cancer cells to bone. Using cortical bone slices, it was shown that exposure of MDA-MB-231 cells to taxoids for 1 h inhibited adhesion to mineralized bone matrices in a dose-dependent manner, with halfmaximal inhibition seen with approximately 80 nmol/l for both taxoids. However, the concentrations of taxoids used to inhibit cell invasion (up to 500 nmol/l) were 25-fold higher than the concentrations required to induce apoptosis. In combination experiments the inhibitory effect of ibandronate on cell adhesion was additive to that of taxoids. Studies done using matrigel invasion assays revealed that exposure of breast cancer cells to ibandronate for 23 h followed by 1 h exposure to taxoids increased inhibitory effects on cell invasion by 70-78% as compared with taxoids alone.

Because breast cancer cell invasion requires both cell migration and digestion of the basement membrane by matrix metalloproteinases (MMPs), bisphosphonates could affect one or the other of these mechanisms. Because ibandronate treatment did not inhibit cell migration in the study of Magnetto et al. [18], it is possible that the bisphosphonate mediates effects on cell invasion by decreasing production of MMPs or by inhibiting their activity. Using similar methods, those investigators determined the order of potency of four bisphosphonates in the inva-

sion assay [19]. The order of potency was found to be as follows: zoledronic acid > ibandronate > NE-10244 (active analogue) > clodronate. The half-maximal inhibition values were found to be <1 pmol/l, 1 pmol/l, 0.5 nmol/l and 50  $\mu$ mol/l, respectively.

NE-58051 had no inhibitory effects on cell invasion [19], which is in accord with its ineffectiveness in preventing cell adhesion, as outlined above. This indicates that the direct inhibitory action of bisphosphonates on breast cancer cells involves the R2 group of the molecule (Fig. 1). On the other hand, NE-10790 (a phosphonocarboxylate analogue of risedronate, in which one of the phosphonate groups is substituted by a carboxyl group) had inhibitory effects on cell invasion to an extent similar to that observed with NE-10244, even though NE-10790 has little effect on antiresorptive activity as compared with NE-10244 on bone. This suggests that the pharmacological mechanism of action of bisphosphonates on tumour cell invasion is distinct from the mechanism of action on bone.

The results from that study [19] also lend support to the suggestion that the inhibitory effects of bisphosphonates on cell invasion are related to the inhibition of the proteolytic activity of MMPs rather than to modulation of their expression. At high concentrations (~100 µmol/l), bisphosphonate treatment inhibited the activity of MMP-2, -9 and -12. Excess of zinc completely reversed bisphosphonateinduced inhibition of cell invasion. In addition, NE-10790 did not inhibit MMP activity. Those findings suggest that the phosphonate groups of bisphosphonates are responsible for the chelation of zinc and the subsequent inhibition of MMP activity. However, although treatment with NE-10790 did not decrease MMP activity, it inhibited breast cancer cell invasion to an extent similar to that observed with NE-10244; this suggests that inhibition of MMP activity is not the sole mechanism by which bisphosphonates inhibit invasion.

# Effects of bisphosphonates on breast cancer cell growth and apoptosis

Previous studies have shown that bisphosphonates reduce metastatic tumour burden in bone with increased apoptosis in osteoclasts [20]. In addition, a number of *in vitro* studies have indicated that bisphosphonate treatment of myeloma cells leads to growth inhibition and induction of apoptosis [21]. Clinical findings have suggested that clodronate treatment may reduce incidence of bone metastases, although these results are not yet conclusive [22]. Taken together, these results indicate that bisphosphonates may exert direct growth inhibitory effects on breast cancer cells, leading to reduced metastatic tumour burden in bone.

This suggestion was confirmed by our group with the demonstration that treatment of cultured breast cancer

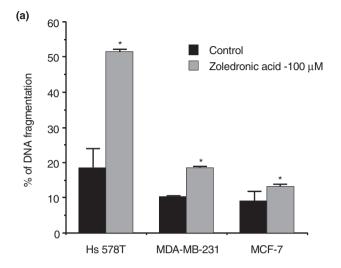
cells with bisphosphonates leads to growth inhibition and induction of apoptosis [23]. In those initial studies, a nonreversible inhibition of cell growth and viability of three human breast cancer cell lines was seen, together with morphological and biochemical changes consistent with apoptosis. Four structurally different bisphosphonates (zoldedronic acid, pamidronate, clodronate and EB-1053) induced apoptosis in a time- and dose-dependent manner. Zoledronic acid was found to be the most potent bisphosphonate, with half-maximal inhibition values in MDA-MB-231 cells of 15 µmol/l; the corresponding values for pamidronate, EB-1053 and clodronate were 40, 1000 and 700 µmol/l, respectively. Furthermore, the order of potency of the bisphosphonates was similar in all three cell lines tested. This finding is in contrast to that of Busch et al. [24], who reported that clodronate is able to reduce survival of MDA-MB-435S but not that of MCF-7 cells.

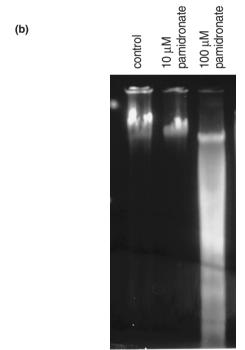
Our findings indicate that the order of bisphosphonate potency on bone resorption is not equivalent to that for inhibiting cell viability in breast cancer cells [23]; it is cell-type-specific. Zoledronic acid has been reported to be 100 times more potent than pamidronate in reducing bone resorption [25], and our study suggests it is approximately threefold more potent than pamidronate in reducing cell viability in breast cancer cells. EB-1053, which is 100 times more potent than pamidronate in inhibiting bone resorption in rats [25], was substantially less effective on breast cancer cells.

We found evidence for fragmentation of chromosomal DNA, a key feature of apoptosis, in MCF-7, MDA-MB-231 and Hs578T breast cancer cells after 2-3 days of treatment with bisphosphonates (Fig. 2) [23]. Induction of apoptosis in MDA-MB-231 cells by pamidronate was accompanied by decreased expression of the antiapoptotic protein bcl-2 as well as cleavage of poly (ADPribose) polymerase, thus implicating the activation of a caspase-dependent pathway. This was later confirmed by Fromigue et al. [26], who showed that inhibition of MCF-7 cell proliferation by four bisphosphonates (zoledronic acid, ibandronate, pamidronate and clodronate) could be abrogated by cotreatment with z-VAD-fmk, a broad-spectrum caspase inhibitor. We similarly found that z-VAD-fmk attenuates loss of MDA-MB-231 breast cancer cell viability in response to pamidronate (Fig. 3) and identified caspase-3 as one of the cell death proteases that are activated by zoledronic acid treatment in MDA-MB-231 cells [27]. Furthermore, Hiraga et al. [28] demonstrated that a selective caspase-3 inhibitor is capable of blocking ibandronate-induced DNA fragmentation in these breast cancer cells.

A recent report by Jagdev et al. [29] presents evidence for synergistic effects of zoledronic acid and paclitaxel on induction of apoptosis in MCF-7 and MDA-MB-231 breast

Figure 2

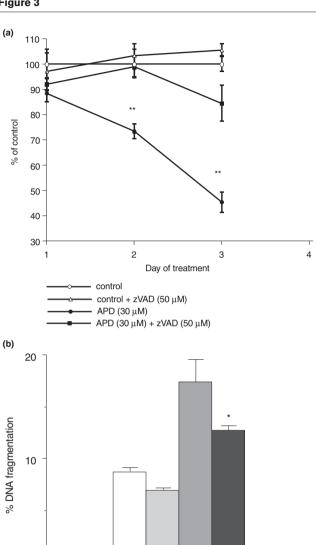


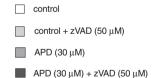


(a) Hs 578T, MDA-MB-231 and MCF-7 breast cancer cells were treated for 2 days with 100 µmol/l zoledronic acid. The percentage of fragmented chromosomal DNA was measured in cells treated with zoledronic acid and compared with that of control (vehicle-treated) cells as previously described [23]. \*P < 0.0005 versus control. (b) Apoptosis was examined by electrophoretic analysis of internucleosomal DNA fragmentation following treatment of MDA-MB-231 cells with 10 and 100 µmol/l pamidronate for 3 days.

cancer cells. Those investigators found a fourfold to fivefold increase in induction of apoptosis in MCF-7 cells when zoledronic acid was used in combination with paclitaxel. This finding is in accord with previous findings of additive effects of bisphosphonates and taxoids on inhibi-

Figure 3





(a) Attenuation by broad-spectrum caspase inhibitor z-VAD-fmk (zVAD) of the effects of pamidronate (APD) on cell viability in MDA-MB-231 cells. Cells were plated in 96-well plates (1 x 10<sup>3</sup> cells/well) and treated with 50  $\mu$ mol/l zVAD 1 h before addition of 30  $\mu$ mol/l APD for 3 days. On days 1, 2 and 3, cell viability was quantitated using MTS dye reduction assay. Results are shown as mean  $\pm$  SD. \*\*P < 0.0001 versus APD + zVAD treatment. (b) Effects of zVAD on APD-induced DNA fragmentation in MDA-MB-231. Cells were plated in 6-well plates at a density of 1  $\times$  10<sup>6</sup>/well and treated with 50  $\mu$ mol/l of zVAD for 1 h before addition of 30 µmol/l APD for 4 days without renewal of medium. Cotreatment with zVAD significantly reduced APD-induced DNA fragmentation. \*P < 0.0005 versus APD treatment alone.

day 4

tion of breast cancer cell invasion and adhesion. In addition. Hiraga et al. [28] reported that bisphosphonates directly induce apoptosis in breast cancer cells that metastasize to bone. Ibandronate (4 µg/mouse per day subcutaneously) was administered after bone metastases had been established by intracardiac inoculation of MDA-MB-231 cells. Inhibition of progression of established osteolytic bone metastases was demonstrated by radiological analysis. Ibandronate significantly decreased tumour burden and increased MDA-MB-231 cell apoptosis in bone metastases. Ibandronate treatment was not able to induce apoptosis in tumours developed by inoculation of MDA-MB-231 cells in the orthotopic mammary fat pads, however, indicating that effects of ibandronate on breast cancer cell apoptosis are restricted to bone in which ibandronate selectively deposits.

The mechanisms by which bisphosphonates promote breast cancer cell apoptosis remain to be established. In osteoclasts and myeloma cells it has been suggested that nitrogen-containing bisphosphonates induce apoptosis by inhibiting enzymes in the mevalonate pathway, preventing the generation of isoprenoid moieties and thereby impairing the isoprenylation (farnesylation and geranylgeranylation) of small GTP proteins such as Ras, Rho and Rac [11,30,31]. Jagdev et al. [29] recently presented evidence that loss of MCF-7 cell viability induced by zoledronic acid could be prevented by coincubation with geranylgeraniol, suggesting a role for impaired protein geranylgeranylation in the effects of the bisphosphonate. The identity of the protein that is affected by zoledronic acid treatment remains to be determined, however. Our own studies have shown that treatment of both MCF-7 and MDA-MB-231 cells with zoledronic acid leads to impaired membrane localization of Ras that is consistent with impaired farnesylation [27]. Further studies are required to determine the signal transduction pathways that are modulated by the alterations in protein isoprenylation induced by bisphosphonates.

#### Conclusion

Laboratory studies increasingly suggest that bisphosphonates can induce important antitumour effects in breast cancer cells *in vitro* by promoting apoptosis, and inhibiting cell adhesion and invasive potential. It is therefore possible that the beneficial effects reported in patients receiving treatment with bisphosphonates may involve direct effects on tumour cells in bone as well as inhibition of osteoclast-mediated bone resorption. It has been noted, however, that the *in vitro* concentrations of bisphosphonates required to induce breast cancer cell apoptosis are higher than those required for osteoclast apoptosis. At the present time the concentrations of bisphosphonates to which tumour cells in bone are exposed are unclear.

Bisphosphonates bind to hydroxyapatite by virtue of their carbon-substituted pyrophosphate structure, and this

accounts for their selective action on the skeleton. The local concentrations of bisphosphonate released from the hydroxyapatite surface into the resorption space are probably considerably higher than the circulating concentration, and have been suggested to approach 800  $\mu$ mol/l [32]. If this is the case then cancer cells may be exposed to concentrations that are sufficient to induce apoptosis.

In summary, the newly recognized direct actions of bisphosphonates on breast cancer cell adhesion, invasion and cell viability indicate that these agents may have a wider role to play in prophylactic treatment of patients suffering from cancers with a propensity to metastasize to bone.

### References

- Elte JWF, Bijovoet OLM, Cleton FJ, van Oosterom AT, Sleeboom HP: Osteolytic metastases in breast carcinoma. Pathogenesis, morbidity and bisphosphonates treatment. Eur J Cancer Oncol 1986, 22:493-500.
- Paterson AHG: Bone metastases in breast cancer, prostate cancer and myeloma. Bone 1987, 8(suppl 1):S17-S22.
- Coleman RE, Rubens RD: The clinical course of bone metastases from breast cancer. Br J Cancer 1987, 55:61-66.
- Mundy GR: Mechanism of osteolytic bone destruction. Bone 1991, 12(suppl 1):S1-S6.
- Lipton A, Theriault RL, Hortobagyi GN, Simeone J, Knight RD, Mellars K, Reitsma DJ, Heffernan M, Seaman JJ: Pamidronate prevents skeletal complications and is effective palliative treatment in women with breast carcinoma and osteolytic bone metastases: long term follow-up of two randomized, placebo-control trials. Cancer 2000, 88:1082-1090.
- Boonekamp PM, van der Wee-Pals LJ, van Wijk-van Lennep MML, Thesing CW, Bijvoet OL: Two modes of action of bisphosphonates on osteoclastic resorption of mineralized matrix. J Bone Miner 1986, 1:27-39.
- Ryzen E, Martodam R, Troxell M, Benson A, Paterson A, Shepard K, Hicks R: Intravenous etidronate in the management of malignant hypercalcaemia. Arch Intern Med 1985, 145:449-452.
- Kanis JA, Urwin GH, Gray RE, Beneton MN, MvCloskey EV, Hamdy NA, Murray SA: Effects of intravenous etidronate on skeletal and calcium metabolism. Am J Med 1987, 82:55-70.
- Lowik CWGM, van der Pluijm G, van der Wee-Pals LJA, Bloys van Trsiong-de Groot H, Bijvoet OLM: Migration and phenotypic transformation of osteoclasts precursors into mature osteoclasts: the effect of a bisphosphonate. J Bone Miner Res 1988, 3:185-192.
- Hughes DE, MacDonald BR, Russell RGG, Gowen M: Inhibition of osteoclast-like cell formation by bisphosphonates in long term cultures of human bone marrow. J Clin Invest 1989, 83: 1930-1935.
- 11. Hughes DE, Wright KR, Uy HL, Sasaki A, Yoneda T, Toodman GD, Mundy GR, Boyce BF: Bisphosphonates promote apoptosis in murine osteoclasts in vitro and in vivo. *J Bone Miner Res* 1995, **10**:1478-1487.
- Selander KS, Monkkonen J, Karhukorpi E, Harkonen P, Hannuniemi R, Vaananen KK: Characteristics of clodronate-induced apoptosis in osteoclasts and macrophages. *Mol Pharmacol* 1996, 50:1127-1138.
- Sasaki A, Boyce BF, Story B, Wright KR, Chapman M, Boyce R, Mundy GR, Yoneda T: Bisphosphonate risedronate reduces metastatic human breast cancer burden in bone in nude mice. Cancer Res 1995, 55:3551-3557.
- Yoneda T, Sasaki A, Dustan C, William PJ, Bauss F, De Clerck YA, Mundy GR: Inhibition of osteolytic bone metastasis of breast cancer by combined treatment with the bisphosphonates ibandronate and tissue inhibitor of the matrix metalloproteinase-2. J Clin Invest 1997, 99:2509-2517.
- Colucci S, Minielli V, Zambonin G, Grno M: Alendronate acts on bone resorption of human osteoclast-like cells through the inhibition of cell attachment to bone surfaces [abstract]. Bone 1995, 17:599.

- van der Pluijm G, Vloedgraven H, van Beek E, van der Wee, Lowik C, Papapoulos SJ: Bisphosphonates inhibit the adhesion of breast cancer cells to bone matrices in vitro. Clin Invest 1996, 98:698-705.
- Boissier S, Magnetto S, Frappart L, Cuzin B, Ebetino FH, Delmas PD, Clezardin P: Bisphosphonates inhibit prostate and breast carcinoma cell adhesion to unmineralized and mineralized bone extracellular matrices. Cancer Res 1997, 57:3890-3894.
- Magnetto S, Boissier S, Delmas PD, Clezardin P: Additive antitumour activities of taxoids in combination with the bisphosphonate ibandronate against invasion and adhesion of human breast carcinoma cells to bone. *Int J Cancer* 1999, 83:263-269.
- Boissier S, Ferreras M, Peyruchaud O, Magnetto S, Ebetino FH, Colombel M, Delmas P, Delaisse J-M, Clezardin P: Bisphosphonates inhibit breast and prostate carcinoma cell invasion, an early event in the formation of bone metastases. Cancer Res 2000, 60:2949-2954.
- Hiraga T, Tanaka S, Yamamoto M, Nakajima T, Ozawa H: Inhibitory effects of bisphosphonate (YM175) on bone resorption induced by a metastatic bone tumour. Bone 1996, 18:1-7.
- Shipman CM, Rogers MJ, Apperley JF, Russell RGG, Croucher Pl: Bisphosphonate induced apoptosis of human myeloma cell lines: a novel antitumour activity. Br J Haematol 1997, 98: 665-672.
- Diel IJ, Solomayer EF, Costa SD, Gollan C, Goerner R, Wallwiener D, Kaufmann M, Bastert G: Reduction in new metastases in breast cancer with adjuvant clodronate treatment. N Engl J Med 1998, 339:357-363.
- Senaratne SG, Pirianov G, Mansi JL, Arnett TR, Colston KW: Bisphosphonates induce apoptosis in human breast cancer cell lines. Br J Cancer 2000, 82:1459-1468.
- Busch M, Rave-Frank M, Hille A, Duhmke E: Influence of clodronate on breast cancer cells in vitro. Eur J Med Res 1998, 3: 427-431.
- 25. Fleisch H: *Bisphosphonates in bone disease*, 3rd ed. New York: Parthenon Publishing Group; 1997.
- Fromigue O, Lagneaux L, Body JJ: Bisphosphonates induce breast cancer cell death in vitro. J Bone Miner Res 2000, 15: 2211-2221.
- 27. Senaratne SG, Colston KW: Mechanisms involved in aminobisphosphonate-induced apoptosis in breast cancer cells [abstract]. Proc Am Assoc Cancer Res 2001, 42:2377.
- Hiraga T, Williams PJ, Mundy GR, Yoneda T: The bisphosphonate lbandronate promotes apoptosis in MDA-MB-231 human breast cancer cells in bone metastases. Cancer Res 2001, 61: 4418-4424.
- Jagdev SP, Coleman RE, Shipman CM, Rostami HA, Croucher PI: The bisphosphonate, zoledronic acid, induces apoptosis of breast cancer cells: evidence for synergy with paclitaxel. Br J Cancer 2001, 84:1126-1134.
- Luckman SP, Coxon FP, Ebetino FH, Russell RGG, Rogers MJ: Heterocycle-containing bisphosphonates causes apoptosis and inhibit bone resorption by preventing protein prenylation: evidence from structure-activity relationships in J774 macrophages. J Bone Miner Res 1998, 13:1668-1678.
- Shipman CM, Croucher PI, Russell RGG, Helfrich MH, Rogers MJ: The bisphosphonates incadronate (YM175) causes apoptosis of human myeloma cells in vitro by inhibiting the mevalonate pathway. Cancer Res 1998, 58:5294-5297.
- Sato M, Gransser W, Endo N, Akins R, Simmons H, Thompson DD, Golub E, Rodan GA: Bisphosphonates action. Alendronate localization in rat bone and effects on osteoclast ultrastructure. J Clin Invest 1991, 88:2095-2105.